

Scientific paper

Atmospheric Solids Analysis Probe with Mass Spectrometry for Chlorpyrifos and Chlorpyrifos-Oxon Determination in Apples

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Abstract

Chlorpyrifos (CPS) is a toxic pesticide present in several pesticide formulations, with low degradability by natural processes. The degradation leads to the toxic metabolite chlorpyrifos-oxon (CPO). The analytical techniques used for the CPS and CPO analysis, like UPLC-PDA and GC-MS, are accurate but also expensive and time consuming, and they need sample pretreatment. In the search of a more rapid and simple analytical procedure, atmospheric solids analysis probe with mass spectrometry (ASAP-MS) was optimized for the determination of CPS and CPO in apples (*Malus domestica* „Idared”). The identification of the analytes was based on protonated ion and isotopic pattern, while the quantification was based on peak intensities. The obtained results were confirmed by re-validated UPLC-PDA and GC-MS techniques. CPS and CPO concentrations determined by ASAP-MS and UPLC-PDA showed moderate discrepancies (on average by 10–20%), thus demonstrating that ASAP-MS can be a semiquantitative tool for the quantification of these compounds. As additional goal of this work, the efficiency of a gamma irradiation treatment to remove CPS and CPO from apples was tested by analyzing their content before and after the irradiation: 89–99% of CPS and CPO were degraded with doses of 3.5–3.8 kGy and 66–72 h of irradiation per sample. Identical degradation results were obtained by UPLC-PDA and ASAP-MS, indicating that the latter technique is well suitable to rapidly check pesticide degradation in apples.

Keywords: GC-MS; UPLC-PDA; ASAP-MS; gamma irradiation

1. Introduction

Chlorpyrifos (O,O-diethyl-O-3,5,6-trichloro-2-pyridylphosphorothioate, CPS, Figure 1) is an organophosphorous insecticide used in agriculture formulations and in the military field. It is a very persistent (resistant to degradation) and non-volatile compound at room temperature. CPS may be detected in fruit, vegetables, water, soil, and body fluids, even months to years after its application.

This occurs especially after unprofessional application of agricultural formulations, like EC (emulsifiable concentrate) and EW (emulsion in water). EC contains 30–50% of active substance, organic solvents (such as high-boiling mineral oils, 40–60%), and emulsifiers, whereas EW contains active substances (about 30%), emulsifier (15%), antifreeze (10%), antifoam, thickener, biocide, stabilizer, buffer, and water. There are also other formulations based on this pesticide.^{1–3}

The degradation process of CPS leads to the formation of chlorpyrifos-oxon (*O,O*-diethyl-*O*-3,5,6-trichloro-2-pyridylphosphate, CPO) and 3,5,6-trichloro-2-pyridinol (TCP). The formation of CPO (Figure 1) occurs in animal liver, where a phosphorus-sulfur bond of CPS is replaced by a phosphorus-oxygen bond.^{4,5}

TCP is non-toxic, but CPS and CPO are very toxic to humans and cause tens of thousands of deaths per year worldwide because of many reasons: type of pesticide application, short waiting period between treatments, higher doses than allowed, and high toxicity. Both CPS and CPO can cause chemical injury leading to serious damages of human hepatocytes (cells of main liver parenchyma), immune, cognitive, and reproductive systems, in the gastrointestinal tract, and to the hormonal status. The most affected patients are children, elderly, and occupationally exposed people.⁶ K. Choi et al. and J. Choi et al. demonstrated that CPO inhibits the esterase in human liver.^{7,8} In case of highly intoxicated liver, metabolic paths change (due to enzyme inhibition) and carcinoma can appear in humans (colorectal carcinoma, liver, lung).⁹ It is known that hepatocytes in humans with polymorphisms do not have the ability to metabolize, detoxify, and inactivate exogenous compounds such as insecticides and drugs.¹⁰ Thus, both European Commission (EU) and U.S. Environmental Protection Agency (EPA) restricted the use of CPS and are planning to forbid it. The CPS and CPO concentrations in different matrices (food, water, agricultural formulation, blood, urine) should be monitored continuously.¹¹

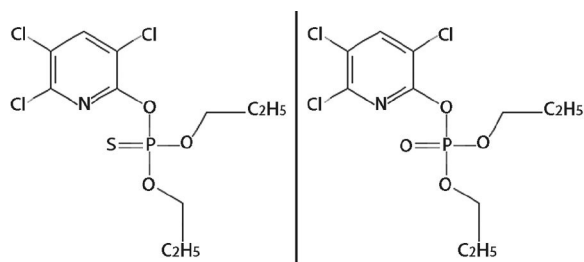


Figure 1. *O,O*-diethyl-*O*-3,5,6-trichloro-2-pyridylphosphorothioate (CPS, left) and *O,O*-diethyl-*O*-3,5,6-trichloro-2-pyridylphosphate (CPO, right).

CPS and CPO concentrations in food and other matrices are most often monitored by chromatographic techniques like high pressure liquid chromatography with photodiode detection (UPLC(HPLC)-PDA), liquid chromatography – mass spectrometry (LC-MS), liquid chromatography – tandem mass spectrometry (LC-MS/MS)¹², gas chromatography with mass spectrometric detector (GC-MS), gas chromatography with nitrogen phosphorus thermoionic detector (GC-NPD), gas chromatography with flame ionization detector (GC-FID),^{13,14} 2D-chromatography with simultaneous analysis by two GC columns (GC×GC-FPD).¹⁵ Other, less used techniques are spectro-

scopic ones like infrared spectroscopy (IR), Fourier transform with infrared spectroscopy (FTIR), and attenuated total reflection – Fourier transform with infrared spectroscopy (ATR-FTIR).¹⁶ Almost all of these techniques are highly expensive, and they require toxic solvents and long time for validation and sample preparation. These techniques are also not suitable to rapidly check the presence or absence of CPS and CPO in food or other matrices.

Atmospheric solids analysis probe with mass spectrometry (ASAP-MS) represents a possible alternative method for the analysis of such analytes.¹⁷ ASAP-MS utilizes heated nitrogen to vaporize the sample and corona discharge to ionize it. This technique is capable of ionizing low polarity compounds, such as CPS and CPO, not amenable to electrospray (ESI), to atmospheric pressure photoionization (APPI), or to atmospheric pressure chemical ionization (APCI), and it is possible to analyze samples via direct introduction at atmospheric pressure, thus shortening the analysis time and simplifying the procedure. In this paper, therefore, an ASAP-MS method was optimized for the rapid and simple determination of CPS and CPO in plant origin food, such as apples. The obtained results were confirmed by values obtained through standard sample preparation methods (extractions) and chromatographic techniques, i.e. gas chromatography with mass spectrometric detection (GC-MS) and reverse-phase ultra-high performance liquid chromatography with photodiode array detector (RP UPLC-PDA), re-validated for these purposes.

Furthermore, as natural processes (sun, air, microbiological) are not effective enough in real time for the removal of CPS from food, forced processes like chemical (acid and base hydrolysis, oxidation by H₂O₂) or physical (gamma irradiation) are necessary for the removal of this pesticide or its conversion into nontoxic compound such as TCP. Therefore, another goal of this work was to evaluate if a gamma irradiation treatment is able to efficiently remove CPS and CPO from apples treated by EC and EW formulations. Analytical measurements were performed with both RP UPLC-PDA and ASAP-MS.

2. Experimental

2.1. Materials

The reference materials for the analysis of CPS (98.0±1.0%) and CPO (98.0±1.0%) were purchased from Dr Ehrenstorfer GmbH (Augsburg, Germany). Multi-residue standard mix with CPS in acetonitrile, Pestanal mix 101 with 20 pesticides and CPS (50 ng/μL, 99.0±0.5%) was purchased from Dr Ehrenstorfer GmbH (Augsburg, Germany). The 30% H₂O₂ solution was purchased from MerckCo (Germany). Radar EC and Radar EW, plant protection formulations (active substance CPS 300 g/L), were purchased from Galenika, Phitopharmacia (Belgrade, Serbia). Apples “Idared” were received from domestic pro-

duction and bought in supermarkets. Methanol, acetonitrile, hexane, petroleum ether, acetone (HPLC grade) were purchased from J. T. Baker (Netherlands). Dichloromethane (HPLC grade) was purchased from Across Organic (New York, USA). Formic acid (98%) was obtained from J. T. Baker (Netherlands). Glacial acetic acid (99.8%, HPLC grade) was a Fisher Chemicals (United Kingdom) product. Deionized water was prepared by Purite Select Fusion System. Multi-walled carbon nanotubes (MWCNT) were a Sigma Aldrich product. QuEChERS kit was composed of two extraction mixtures: a mixture of 1.5 g CH_3COONa and 6 g MgSO_4 , and a PSA sorbent. Cartridges used for analytes separation were Oasis® HLB (hydrophilic-lipophilic balance) from Waters (Milford, MA, USA).

2. 2. Solutions

Standard stock solutions of CPS and CPO were prepared by dissolving 10 mg of CPS and 5 mg of CPO in 1000 μL of methanol (or dichloromethane, depending on the instrumental technique used), and then put in a volumetric Eppendorf tube. The stock solutions were diluted in hexane or in deionized water (pH 6.8) to prepare the desired concentrations for the calibration curves for GC-MS or UPLC-PDA, respectively.

EW and EC formulations stock solutions were prepared by dissolving 100 μL of formulation in a 10 mL volumetric flask with dichloromethane (or in methanol/water mixture 1:3, v/v), at a concentration of 3000 mg/L calculated on active component, and stored at -18°C when not in use.

2. 3. Instrumentations and Methods

2. 3. 1. ASAP-MS

ASAP-MS with triple quadrupole analyzer (TQD, Acquity, Waters, USA) was used for the mass spectrometry analysis of CPS and CPO. The ASAP compartment consists of set probes and removable sample insertions, corona pin, and glass sample capillaries. The capillary glass tubes were sealed at the end and baked at 500°C to remove possible contaminations. The whole assembly (100 mm in length) was inserted in the mass spectrometer. Positive ions were recorded between 50 and 700 Da. The ASAP probe with sample was inserted into the sealed source enclosure and desolvation gas was rapidly heated to 300°C . The heated nitrogen volatilizes the sample from a glass capillary tip, which is then ionized with a corona discharge pin, and MS spectrum is generated. The mass accuracy of the data was based on the instrument internal calibration.

Instrumental parameters were varied to avoid CPS and CPO fragmentation. Values accepted after optimization were: corona voltage 3.8 kV, extraction cone 2.0 V, source temperature 150°C , desolvation gas flow 650 L h^{-1} , sampling cone voltage 35 V, desolvation temperature 300°C . Data were processed by the MassLynx software, version 4.1.

2. 3. 2. GC-MS

The GC-MS QP2010 Ultra instrument (Shimadzu, Kyoto, Japan) was used. A Rtx*-1 (RESTEK, Crossbond® 100% dimethylpolysiloxane, $30\text{ m} \times 0.25\text{ mm I.D.}$, $0.25\text{ }\mu\text{m}$ film thickness) column was used. The injector temperature was 290°C . The GC temperature program was held at 50°C for 1 min, then elevated to 140°C at rate of 20°C/min , and to 300°C at rate 10°C/min , with a hold time of 6 min. Helium was used as the carrier gas at approximately 1.0 mL/min pulsed in splitless mode. The mass spectrometric detector was operated in an electron impact ionization mode with an ionizing energy of 70 eV and scanning range from 50 to 500 m/z .

All measurements have been done in triplicate. Quantification of CPS and CPO has been done by external calibration curves. The calibration ranges of CPS and CPO explored in this work were 0.2–5 mg/L and 0.3–3.0 mg/L, respectively; linearity in the signal/concentration ratio was obtained also up to 30 mg/L for CPS and 25 mg/L for CPO. The correlation coefficient R^2 was higher than 0.99. The limit of detection (LOD) for CPS was 0.08 mg/L and it was 0.1 mg/L for CPO, calculated for samples injected three times. The limit of quantification (LOQ) was 0.2 mg/L for CPS and 0.3 mg/L for CPO. Relative standard deviations (RSD) were determined from ten measurements of a CPS standard solution ($C = 2.5\text{ mg/L}$) and a CPO standard solution ($C = 1.5\text{ mg/L}$). The RSD was 4.1% for CPS and 4.8% for CPO.

2. 3. 3. RP-UPLC-PDA

Reversed-phase ultra high performance liquid chromatography (RP UPLC) was performed with an Acquity system with PDA detector (Waters, USA). Samples were injected automatically; the volume of injected sample was 10 μL . Acquity BEH Reverse Phase C18 ($1.7\text{ }\mu\text{m} \times 100\text{ mm} \times 2.1\text{ mm}$) column was used as a stationary phase for chromatographic separations. Mobile phases contained 0.1% formic acid in water (v/v) (A) and 0.1% formic acid in acetonitrile (v/v) (B). The following gradient elution conditions were used: 0–1.5 min 45% A; 1.5–2 min 40% B; 2–2.5 min 20% B; 2.5–4.5 min 2% B; 4.5–4.8 min 55% B; 4.8–6.0 min 50% B. Flow rate was set at 1 mL/min , and column temperature was 30°C . The chosen absorption wavelengths were 230 and 280 nm. Quantification of CPS and CPO has been done by external calibration curves. All measurements have been done in triplicate.

The calibration ranges of CPS and CPO were 0.1–5 mg/L and 0.03–2.5 mg/L, respectively; linearity in the signal/concentration ratio was obtained also up to 30 mg/L for CPS and 20 mg/L for CPO. The correlation coefficient R^2 was higher than 0.99. The LOD for CPS was 0.03 mg/L and it was 0.01 mg/L for CPO, calculated for samples injected three times. The LOQ was 0.10 mg/L for CPS and 0.03 mg/L for CPO. The accuracy and precision were evaluated by means of the recovery: values obtained by

post-extraction addition of standard to treated apples were 98–108% and 96–104% for the two compounds, respectively. RSD values were determined from ten measurements of CPS standard solution ($C = 0.3$ mg/L) and CPO standard solution ($C = 0.8$ mg/L). The RSD was 2.2% for CPS and 3.3% for CPO.

2. 4. Gamma Irradiation Experiments

Gamma irradiation experiments were carried out in the metrology dosimetry laboratory at the Institute of Nuclear Sciences (Belgrade). The radiation unit IRPIK-B was used as a functional generator with a source of gamma radiation Co-60. The measured value of the absorbed dose in the air at the reference point for the irradiation was 48.27 Gy/h. At the point at which samples were positioned, the absorbed dose in water was 53.51 Gy/h. After each individual campaign irradiation, the total value of the absorbed dose in each sample was 3.5–3.8 kGy, depending on time of irradiation. In general, the relation between the absorbed dose in air and absorbed dose in water as the material that is irradiated with gamma rays is:

$$D_W = D_{air} \cdot \frac{\left(\frac{\mu}{\rho}\right)_{en,air}}{\left(\frac{\mu}{\rho}\right)_{en,W}} \quad (1)$$

where D_W and D_{air} are the absorbed dose in the water and

absorbed dose in the air, respectively, and $\left(\frac{\mu}{\rho}\right)_{en,air}$ and $\left(\frac{\mu}{\rho}\right)_{en,W}$

are the energy absorption coefficients for air and water, respectively. It was assumed that absorption of irradiation in apple samples is equivalent to that in water.

2. 5. Sample Preparation

The plant protection formulations based on CPS were applied to 20 samples of apples “Idared” bought in supermarkets, and to 20 samples of the same variety obtained from orchards. The supermarket samples were sprayed by 2 mL of EC formulation (dissolved in a water methanol mixture, 3:1, v/v) under 100 kPa air pressure from a Potter precision laboratory spray tower (Burkard Scientific, UK). Spraying was performed under controlled conditions of temperature (17 °C) and relative humidity (45%). The treated samples were collected in plastic bags after treatment and stored at –18 °C until further analysis. Some apples were further treated with 30% H₂O₂ solution to cause initial degradation of EC formulation to form CPO.

For ASAP-MS analysis, the samples were brought to room temperature. Apples were peeled, and their skin was cut into small pieces (about 1×1 cm), and the ASAP-MS

probe was dipped into the apple pieces. The skin was weighted before and after dipping the ASAP-MS probe. The sample was then inserted into the ionization source chamber. No particular sample pretreatment was needed for the ASAP-MS analysis, and MS data for each sample were collected in few seconds.

For the chromatographic analysis, apples were homogenized (15 g in 30 mL acetone), vortexed and centrifuged for 10 min at 3500 rpm. The procedure was repeated three times, and acetone extract was collected. The remaining apple homogenate was further mixed with 20 mL of petrol ether and 10 mL of dichloromethane. After centrifugation, the upper organic layer was collected and added to acetone extract by this modified Luke procedure.¹⁸ Combined extracts were evaporated on a water bath at 42–62 °C and reconstituted with methanol for UPLC-PDA analysis or dichloromethane for GC-MS analysis. For some apples, the skin was separated from the mesoderm, and each sample was treated separately.

Three extraction techniques, other than the described Luke procedure, have been tested: liquid phase extraction followed by analytes separation in Oasis® HLB cartridge, extraction with multiwall carbon nanotubes (MWCNT) and extraction with QuEChERS kit. For the liquid phase extraction, 5 g of apples have been homogenized and extracted with 20 mL of acetonitrile; 5 mL of extract were diluted to 100 mL with a 50:50 water/methanol mixture, and eluted through the Oasis® HLB cartridge (previously conditioned with methanol). Eluted solutions were evaporated to dryness at 42–64 °C, and reconstituted with methanol for UPLC-PDA analysis or dichloromethane for GC-MS analysis. For the extraction with MWCNT, 6 mL PTFE tubes (Strata) were filled with 0.035 g MWCNT, sample was added, and pH was then set at 6–7.5 to enhance the analyte extraction. Analytes were eluted with a methanol solution, and then samples were evaporated to dryness and reconstituted as described above for the liquid phase extraction. As regards the QuEChERS kit extraction, 15 g of homogenized sample were loaded into an empty 50 mL tube, and 15 mL of a 1% CH₃COOH in acetonitrile solution were added. The first QuEChERS mixture was then added. After centrifugation at 1500 rpm for 5 min, 3 mL of supernatant solution were transferred into another tube containing the second QuEChERS mixture. After centrifugation at 3000 rpm for 5 min, 100 µL of the final extract were transferred into a vial and diluted with methanol.

The extraction efficiency η (%) was calculated for each method. The combined Luke (Dutch) procedure gave an efficiency of 94% ± 8%, which was the highest among all obtained efficiencies. For the liquid phase extraction it was 84% ± 13%, for the MWCNT it was 88% ± 9%, and for the QuEChERS kit it was 90% ± 8%. Therefore, the combined Luke (Dutch) procedure was chosen for apple preparation for the UPLC-PDA analyses.

3. Results and Discussion

3.1. ASAP-MS

The interpretation of ASAP-MS spectra of CPS and CPO was performed in accordance to m/z values and isotopic pattern. The protonated CPS is at m/z 351.5 ± 0.5 Da, and the protonated CPO is at m/z 335.5 ± 0.5 Da. The non-toxic metabolite TCP can also be observed at m/z 198 with very low abundance. CPS and CPO are chlorinated compounds with three chlorine atoms which are excellently suitable for interpretation by isotope distribution. For example, the isotopic pattern for protonated CPS is 350:351:352:353:354:355:356, and calculated relative ion intensities are 97:11:100:11:36:4:5, respectively. In this pattern the even m/z values (350, 352...) stem mainly from the chlorine isotopes ^{35}Cl (75.8% natural abundance) and ^{37}Cl (24.2% natural abundance), and the uneven values (351, 353...) from the carbon isotopes ^{12}C (98.9% natural abundance) and ^{13}C (1.1% natural abundance). Theoretical ratio is in good agreement with ion ratio detected experimentally from standard and from apple samples (an example of experimental pattern is reported in Figure 1S in the Appendix A – Supporting Information).^{7,19}

The ASAP-MS spectra of a blank apple and of an apple treated by 50 mg/L EC formulation and with H_2O_2 are shown in Figure 2. If the two spectra are compared, the appearance of two new intense signals (10-fold more intense than other m/z signals) due to CPS and CPO can be observed: CPS was added with the EC formulation, whereas CPO was produced by the H_2O_2 addition which simulates CPS degradation.

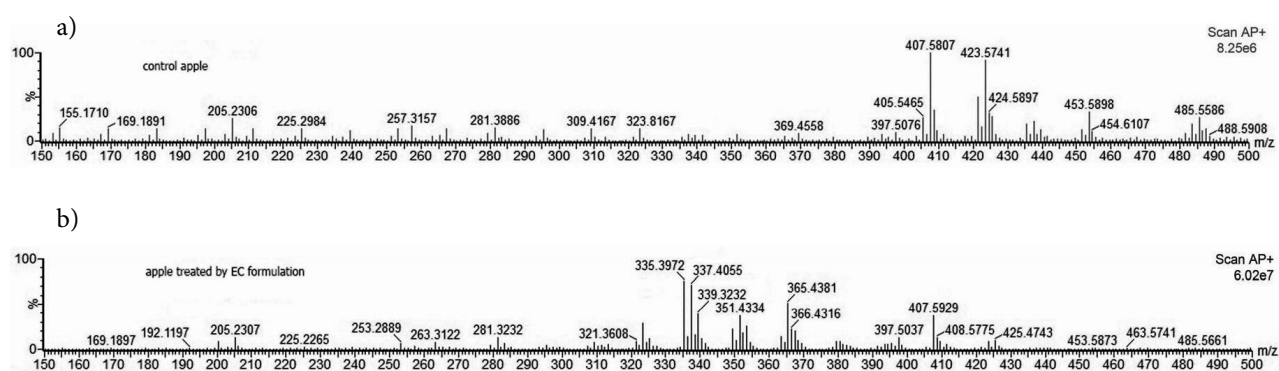


Figure 2. ASAP-MS spectra of a control apple (a) and of an apple treated with 50 mg/L EC formulation and with H_2O_2 (b).

According to literature data,^{20–22} some additional compounds can be likely identified in Figure 2: naringenin pentose (m/z 405), *p*-coumaric pentose (m/z 326), phloridzin (m/z 435), hyperoside (quercetin 3-*O*-galactoside) or isoquercetin (m/z 463), ferulic acid (m/z 368), cyanidin pentosine (m/z 419), cyanidin hexoside (m/z 449), and quercetin pentoside (m/z 435). In principle, therefore,

ASAP-MS might be used also for the detection of these additional compounds.

3.2. Results of Chromatographic Analyses

Chromatographic techniques (GC-MS and UPLC-PDA) were validated and applied for accurate measurements of CPO and CPS in apples, in order to acquire the “true” concentrations of these compounds.

The identity of CPS and CPO in GC-MS has been confirmed from the electron impact (EI) spectra which gave confirmatory product ions, as shown in the Supporting Information (Figures 2S and 3S). The same ions are reported in Wiley database libraries. As well, the retention times of CPS and CPO were identical to those of the standard materials. The identification by UPLC-PDA techniques has been done by the retention time, which was identical to that of the standard material. The retention times were 4.23 min for CPS, 3.08 min for CPO, and 2.01 min for TCP.

From the validation data reported in the Experimental section, UPLC-PDA demonstrated to have slightly better analytical performances (LOD, LOQ, recovery) than GC-MS. For this reason, the subsequent quantitative analyses have been performed using UPLC-PDA. Some quantitative results obtained on apples (blank or treated with pesticide formulations) are shown in Table 1S in the Supporting Information. These results show that CPS residues remained on apple skin, while apple mesoderm did not contain the pesticide. Untreated samples (domestic apple) were also CPS free.

3.3. Comparison Between ASAP-MS and Chromatographic Measurement of CPS and CPO in Apples

The results obtained in the analysis of two samples for both CPS and CPO by using two different techniques, UPLC-PDA and ASAP-MS, are reported in Table 1. The first column reports the concentrations of EC or EW formulations used for apples spraying.

Although results obtained by ASAP-MS method were different than those obtained by UPLC-PDA meth-

od, the order of magnitude of the values given by the two techniques was the same, and relative differences were not very large (around 10–20%). Indeed, ASAP-MS results were obtained in few minutes, whereas UPLC-PDA values required a 10–100-fold longer time. This indicated that ASAP-MS technique can represent a rapid and semiquantitative method for the analysis of CPS and CPO.

The quantitative results for the measure of the degradation efficiency were obtained with UPLC-PDA and are shown in Table 2. The last column reports the removal efficiency of CPS (if applicable also of CPO), as a function of the gamma irradiation doses reported in the previous column and/or of the H₂O₂ pretreatment.

The pretreatment with H₂O₂ caused only a limited

Table 1. Comparison between results from UPLC-PDA and from ASAP-MS in the analysis of two apple samples.

sample	technique	CPS [mg/kg]	CPO [mg/kg]
treated with 50 – EC	UPLC-PDA	0.65±0.02	0.06±0.01
treated with 50 – EC	ASAP-MS	0.58±0.03	0.05±0.01
treated with 50 – EW	UPLC-PDA	0.60±0.03	0.26±0.02
treated with 50 – EW	ASAP-MS	0.54±0.02	0.21±0.02

3. 4. Effect of Gamma Ray Treatment on the Removal of CPS and CPO from Treated Apples

The GC-MS chromatogram (recorded as total ion chromatogram, TIC) obtained in EC formulations (5 mg/L) before gamma irradiation is presented in Figure 4Sa in the Supporting Information. The peak of CPS was observed at $t_R = 18.27$ min. Other minor formulation ingredients can be seen at different t_R values. When the analysis was repeated after gamma irradiation, the CPS peak almost disappeared (Figure 4Sb): CPS peak area is around 10–15% of the same peak in Figure 4Sa. Gamma irradiation caused the appearance of the CPO peak at $t_R = 18.03$ min and of TCP at $t_R = 12.77$ min. Both compounds were produced from the degradation of CPS.

Figure 3 shows the ASAP-MS spectra of a blank apple gamma irradiated (the same sample of Figure 2a), and of an apple treated with 50 mg/L EC formulation, with H₂O₂ and then gamma irradiated (the same sample of Figure 2b). Both apples show an almost identical spectrum, indicating that the degradation was effective, and that the treated apple was almost identical to a blank (i.e. not polluted) apple.

degradation of CPS (around 35–40%), and it caused the production of large amounts of CPO due to chemical oxidation of CPS. The exposure to gamma rays caused the removal of CPS and of CPO depending on the irradiation dose and on the irradiation time. 2.5 kGy for 66 h was not sufficient for pesticide removal, as the removal percentage was below 50%. A good efficiency was achieved for a 3.5 kGy dose and for a 66 h treatment, as the removal percentage of CPS reached 90%. The upper dose of 3.8 kGy for 72 h gave even better results, but the increase with respect to 3.5 kGy for 66 h was not very large. Little amounts of CPO appear as a degradation product of CPS after gamma irradiation. However, if CPO is present on apples in large amounts, it is degraded as well by gamma irradiation, as demonstrated in samples pretreated with H₂O₂. The parameters of gamma irradiation and their effects are in agreement with literature data for interaction of gamma irradiation with water medium.^{23–25}

The removal efficiency of gamma irradiation can be easily checked also by ASAP-MS, by measuring the absolute intensities of the relevant pesticide peaks before and after irradiation. For example, with reference to Figures 2 and 3, in apples treated with 50 – EC the average intensi-

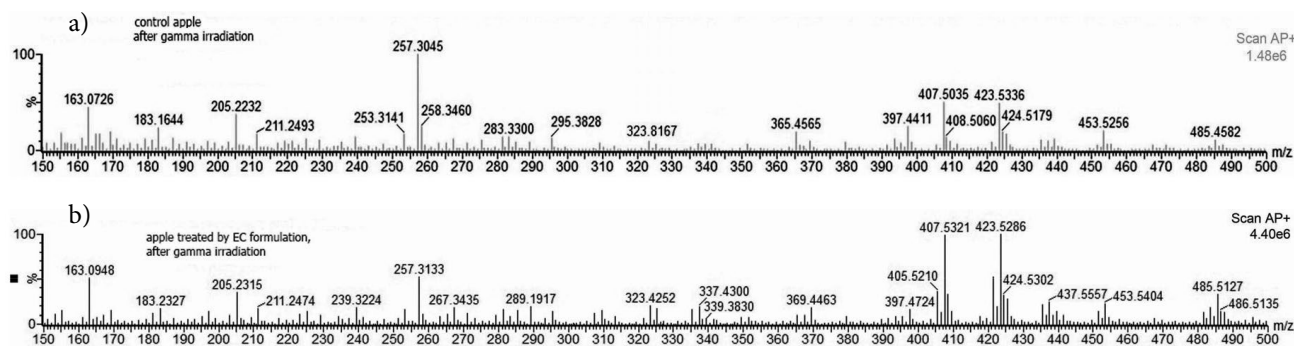


Figure 3. ASAP-MS spectra of a control apple after gamma irradiation (a) and of an apple treated with 50 mg/L EC formulation and with H₂O₂ and then gamma irradiated (b).

Table 2. Removal of CPS and CPO by gamma ray irradiation and, if used, by H₂O₂ pre-treatment. Values were determined by UPLC-PDA.

sample	CPS [mg/kg]	CPO [mg/kg]	H ₂ O ₂ pretr.	Gamma irradi. doses [kGy]/time[h]	Removal of CPS or CPO [%]
Blank apple	≤0.03	≤0.03	no	–	–
treated with 5 – EC	4.93±0.07	–	no	–	–
treated with 5 – EC	2.76±0.04	0.14±0.02	no	2.5/66	44
treated with 5 – EC	0.55±0.02	0.03±0.01	no	3.5/66	89
treated with 5 – EC	0.56±0.02	0.03±0.01	no	3.8/72	88
treated with 5 – EW	4.96±0.05	–	no	–	–
treated with 5 – EW	2.72±0.07	0.13±0.01	no	2.5/66	45
treated with 5 – EW	0.54±0.02	0.03±0.01	no	3.8/72	89
treated with 50 – EC	47.2±1.0	–	no	–	–
treated with 50 – EC	2.7±0.1	0.18±0.02	no	3.8/72	94
treated with 50 – EC	30.2±1.0	14.10±0.08	yes	–	36
treated with 50 – EC	0.65±0.02	0.06±0.01	yes	3.8/72	98 CPS/99 CPO
treated with 50 – EW	48.2±1.1	–	no	–	–
treated with 50 – EW	2.6±0.1	0.16±0.02	no	3.8/72	94
treated with 50 – EW	29.2±1.1	16.2±0.1	yes	–	39
treated with 50 – EW	0.60±0.03	0.26±0.02	yes	3.8/72	98 CPS/98 CPO

ties of the protonated CPS ion, $m/z=351$, before irradiation were $373 \cdot 10^7$. The intensities of the same ion after gamma irradiation were $4.4 \cdot 10^7$. The efficiency of CPS removal can be thus calculated to be 99%, in very good agreement with the value (98%) determined by UPLC-PDA. Also, the average intensities of the protonated CPO ion at $m/z = 335$ was $259 \cdot 10^7$ before irradiation and $3.5 \cdot 10^7$ after irradiation, so that the effect of removal of CPO was 99%, identical to what was found by UPLC-PDA. ASAP-MS can therefore represent a rapid tool to accurately check CPS and CPO removal from apples.

4. Conclusions

ASAP-MS was optimized for the determination (identification and semiquantification) of CPS and CPO in raw apples. The technique is simple, rapid and economical, as it enables the analysis without purification and preparation steps. Results are indeed not very accurate if compared with those of validated chromatographic techniques (bias was 10–20%), thus indicating that ASAP-MS represents a semiquantitative tool, but these values can be obtained much more rapidly than with validated chromatographic methods.

In the frame of checking possible degradation treatments of CPS and CPO in apples, gamma irradiation demonstrated to be effective enough with a 3.5 kGy dose for a 66 h treatment. ASAP-MS gave in this case accurate degradation percentages by simply measuring the absolute percentages detected for the CPS and CPO ions before and after irradiation. Degradation percentages obtained from ASAP-MS were in very good agreement with the ones obtained by chromatography. This indicates that ASAP-MS can be very well suitable to rapidly and easily check CPS and CPO removal from apples.

5. Acknowledgements

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Appendix A: Supporting Information

5. References

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Povzetek

Klorpirifos (CPS) je strupen pesticid, ki se nahaja v številnih fitofarmacevtskih sredstvih in se pod naravnimi pogoji le malo razgrajuje. Razkroj vodi do nastanka strupenega metabolita klorpirifos-oksona (CPO). Analizne tehnike za določevanje CPS in CPO, kot sta UPLC-PDA in GC-MS, so točne, vendar tudi drage in časovno zamudne ter potrebujejo predpripravo vzorca. Z namenom najti hitrejši in bolj preprost analizni postopek smo za določitev CPS in CPO v jabolkih (*Malus domestica* „Idared”) optimizirali sondo za trdne vzorce pri atmosferskem tlaku v povezavi z masno spektrometrijo (ASAP-MS). Identifikacijo analitov smo izvedli na osnovi protoniranega iona in izotopskega vzorca, medtem ko je bila kvantifikacija osnovana na intenziteti vrhov. Dobljene rezultate smo potrdili z revalidiranimi UPLC-PDA in GC-MS metodama. Koncentracije CPS in CPO, določene z ASAP-MS in UPLC-PDA, so se nekoliko razlikovale (v povprečju za 10–20%), kar je pokazalo, da je ASAP-MS lahko semikvantitativno orodje za kvantifikacijo teh spojin. Dodaten namen te raziskave je bil preizkusiti učinkovitost gama ožarčenja za odstranjevanje CPS in CPO iz jabolk, zato smo jih analizirali pred in po ožarčenju: 89–99% CPS in CPO se je razgradilo pri dozah 3,5–3,8 kGy in času ožarčenja 66–72 h na vzorec. Z UPLC-PDA in ASAP-MS smo dobili identične rezultate razgradnje, kar pomeni, da je slednja tehnika zelo primerna za hitro preverjanje razgradnje pesticidov v jabolkih.