

## Research Article

# Synthesis, Characterization, and Antioxidant Activities of Genistein, Biochanin A, and Their Analogues

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Received 16 June 2017; Accepted 28 September 2017; Published 16 January 2018

Academic Editor: Maria B. P. P. Oliveira

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A series of naturally occurring genistein (**3**) and biochanin A (**4**) compounds and their analogues were synthesized from phloroglucinol. The structures of all the synthesized compounds were established by the combined use of <sup>1</sup>H NMR, <sup>13</sup>C NMR, IR spectral data, and mass spectrometry; their antioxidant activities were investigated. Most of the synthesized compounds show moderate-to-high activity; only two compounds exhibit no significant activity.

## 1. Introduction

Isoflavonoids (**2**) are one of the main subclasses of flavonoids (**1**) which contain C6-C3-C6 carbon skeleton based on 3-phenylchroman [1]. They are widely distributed in many plants [2, 3] and in various common foods which are particularly abundant in seeds and other parts of *leguminous plants* [4, 5]. Significant amounts of isoflavonoids were found in soybeans [6]. Epidemiological studies show that a high consumption of soybean derived foods correlates to a low incidence of hormone related diseases such as cancers, osteoporosis, and cardiovascular diseases [7]. Genistein (**3**) has been one of the most widely studied natural products and has exhibited a wide range of biological activities such as anticancer [8–11], antioxidant [12–16], and antimicrobial activities [17, 18]. Genistein can bind to both the Estrogen Receptor alpha (ER $\alpha$ ) and the Estrogen Receptor beta (ER $\beta$ ), although it has a higher affinity for the ER $\beta$  [19], and genistein is thought to exert its estrogenic effects through mechanisms similar to those of estradiol [20]. Biochanin A (**4**) is the 4'-O-methyl derivative of genistein and biochanin A is the predominant isoflavone found in *alfalfa*, *Trifolium pratense*, and *Cicer arietinum* [21] which has an inhibitory and apoptogenic effect on certain cancer cells such as pancreatic cancer and prostate cancer [22, 23].

The biological and biochemical activity, potential chemopreventive property, therapeutic properties, and genistein affinity towards a large variety of molecular targets attracted the interest of many researchers. The number of publications regarding the synthesis and biological evaluation of genistein and its derivatives has increased [24–27]. This paper presents the synthesis of genistein, biochanin A, and their synthetic analogues to investigate the antioxidant activities and the effect of various substituents on the activity of the molecule.

## 2. Results and Discussion

### 2.1. Antioxidant Activity

**2.1.1. DPPH (1,1-Diphenyl-2-picrylhydrazyl) Radical Scavenging Activity.** The DPPH radical scavenging activity of the synthesized compounds was carried out according to the method of [28]. The *in vitro* antioxidant activity of the final compounds **4(a–h)** was evaluated by DPPH (1,1-diphenyl-2-picrylhydrazyl) radical scavenging method. Ascorbic acid was used as the positive control. The DPPH radical scavenging activity was carried out at concentration of 100  $\mu$ g/ml and the results were reported as average of three replicates. When DPPH reacts with an antioxidant compound, the decrease in absorbance observed because of the reaction

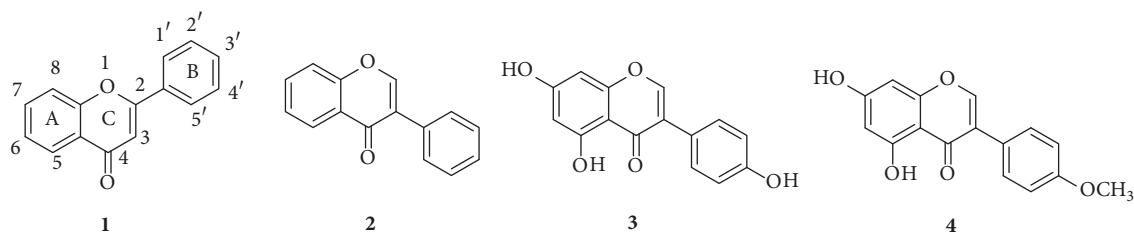


FIGURE 1: Structure of flavonoid (1), isoflavonoid (2), genistein (3), and biochanin A (4).

between antioxidant molecules and radical progresses, which results in the scavenging of the radical by hydrogen donation. The hydrogen donating activity measured at 517 nm showed a significant relationship between concentration of compounds and percentage of inhibition. The structures of flavonoid, isoflavonoid, genistein, and biochanin A are shown in Figure 1.

The DPPH radical scavenging activity was expressed as % of inhibition and results are tabulated in Table 1. Among the target compounds **4(a–h)**, the compounds **4c** (86.95) and **4b** (84.89) exhibited maximum DPPH free radical scavenging activity followed by the compounds **4g** (76.36), **4e** (74.77), **4f** (73.81), and **4h** (65.56) which possess good radical scavenging activity when compared with the reference standard ascorbic acid (92.22). The compounds **4a** and **4d** exhibited no significant radical scavenging activity.

**2.2. Procedure.** The synthesized compounds were dissolved in DMSO at a concentration of 100  $\mu\text{g/ml}$ . Ascorbic acid was used as a reference standard. 0.004% of DPPH was freshly prepared in methanol. 2 ml of DPPH was added to each test tube containing 100  $\mu\text{g/ml}$  concentration of synthesized compounds **4(a–h)** (1 ml) and of standard solution (1 ml). It was shaken vigorously. They were then allowed to stand for 30 min at room temperature in dark place. The control was carried without addition of DPPH and the synthesized compounds. DMSO was used for base line corrections and absorbance (OD) of sample was measured at 517 nm. The following formula was used to interpret the value of the sample:

$$\begin{aligned} &\% \text{ Radical Scavenging Activity} \\ &= \left[ \frac{(\text{control OD} - \text{sample OD})}{\text{control O.D}} \right] \times 100. \end{aligned} \quad (1)$$

**2.3. Conclusions.** The aim of the present work was to compare the radical scavenging activities of genistein and its analogues, which have various substituents at 4' position of B-ring, and study the effect of these substituents on antioxidant activity.

The maximum inhibitory effect was exhibited by compound **4c**, (5,7-dihydroxy-3-phenyl-4*H*-chromen-4-one) which lacks hydroxyl substituent on the B-ring, it has two hydroxyl groups on the A-ring (at C-5 and C-7), and the potent activity suggested that, in cases where the B-ring could not contribute to the inhibitory activity of the isoflavonoids,

TABLE 1: DPPH radical scavenging activity of the target compounds.

Entry	Compound	% inhibition at 100 $\mu\text{g/ml}$ <sup>a</sup>
(1)	Biochanin A (4a)	>100
(2)	Genistein (4b)	84.89 $\pm$ 2.155
(3)	4c	86.95 $\pm$ 3.94
(4)	4d	>100
(5)	4e	74.77 $\pm$ 1.90
(6)	4f	73.81 $\pm$ 1.61
(7)	4g	76.36 $\pm$ 1.42
(8)	4h	65.56 $\pm$ 1.15
(9)	Ascorbic acid	92.22 $\pm$ 2.91

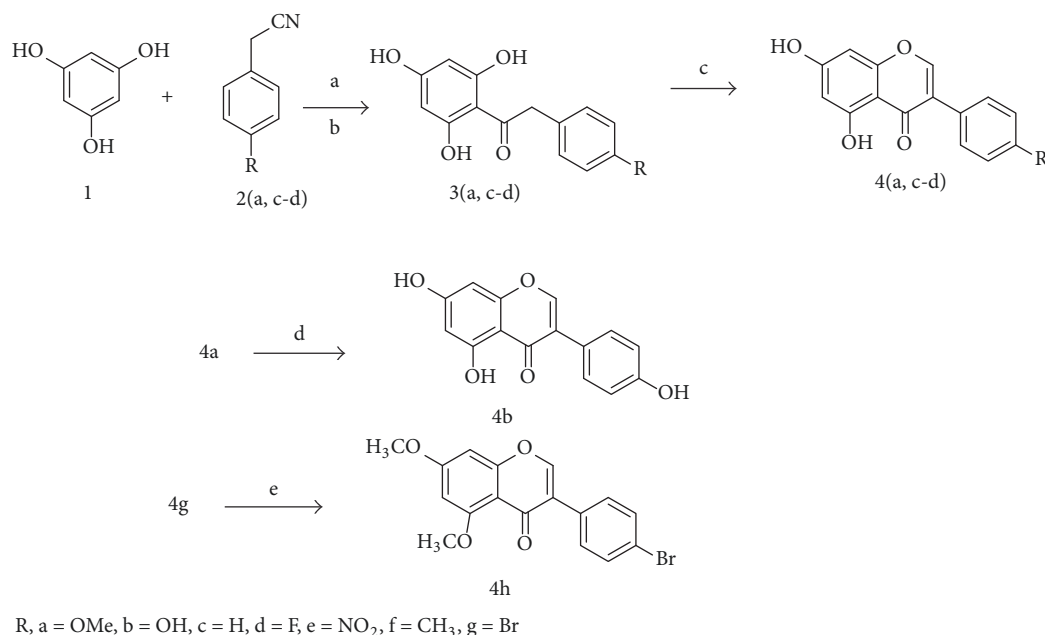
<sup>a</sup>Each value represents mean  $\pm$  SD of three independent experiments.

the hydroxyl group on the A-ring may become a major factor to show the activity. Compound **4h** which has no hydroxyl group exhibited radical scavenging activity (65.56  $\pm$  1.15). Compounds **4e**, **4f**, and **4g** have comparable inhibitory activities. **4g** (76.36  $\pm$  1.42), **4e** (74.77  $\pm$  1.90), and **4f** (73.81  $\pm$  1.61) exhibited moderate activity. The remaining two compounds, **4a** and **4d**, exhibited no significant radical scavenging activity.

### 3. Experimental Section

**3.1. Materials and Methods.** All chemicals and solvents were purchased from Sigma-Aldrich and used without further purification; the reaction process was monitored by TLC silica gel plates; the purification of the products was performed using column chromatography using silica gel (100–200 mesh). Melting points were measured in open capillary tubes and were uncorrected; infrared (IR) spectra were recorded using FT-IR Bruker Alpha spectrometer. NMR spectra were recorded on Bruker (400 MHz) spectrometer using TMS as the internal standard; mass spectra were recorded on an Agilent 110 Lc/MSD. Elemental analyses were performed on a Vario EL-III. The antioxidant activity of all the target compounds (**4a–h**) was investigated using DPPH (1,1-diphenyl-2-picrylhydrazyl) radical method.

**3.2. Chemistry.** The titled compounds (**4a–h**) described in this study were prepared as outlined in Scheme 1, according to the literature method [29]. The intermediate, substituted trihydroxybenzoin (**3a**, **3c–g**) was prepared by acylation of phloroglucinol (**1**) with appropriately substituted phenyl acetonitrile (**2a**, **2c–g**), catalyzed by HCl gas and anhydrous



SCHEME 1: Reagent and condition: (a) HCl<sub>(gas)</sub>, anhydrous ZnCl<sub>2</sub>, dry Et<sub>2</sub>O, 0°C; (b) H<sub>2</sub>O, reflux; (c) Et<sub>2</sub>O·BF<sub>3</sub>, DMF, POCl<sub>3</sub>, 60–70°C, 4 h; (d) AlCl<sub>3</sub>, toluene, 140°C, 6 h; (e) DMS, K<sub>2</sub>CO<sub>3</sub>, Me<sub>2</sub>CO, 60°C, 2 h.

ZnCl<sub>2</sub> in dry ether. Cyclization of the intermediate (**3a**, **3c–g**) with reagents of Et<sub>2</sub>O·BF<sub>3</sub> and DMF/POCl<sub>3</sub> yielded the desired substituted isoflavonoids in good yield. There are several reagents for the demethylation of methyl aryl ethers; we used anhydrous AlCl<sub>3</sub> to synthesize genistein (**4b**) from biochanin A (**4a**) by demethylation of 4'-O-methyl of biochanin A. **4h** was prepared by methylation of **4g** using dimethyl sulphate in acetone in the presence of K<sub>2</sub>CO<sub>3</sub>.

### 3.3. Synthesis and Characterization

**3.3.1. General Procedure for the Synthesis of Substituted 2',4',6'-Trihydroxydeoxybenzoins (3a, 3c–g).** To a solution of phloroglucinol (**1**) (5 g, 0.039 mol) and substituted phenyl acetonitrile (**2a**, **2c–g**) (0.044 mol), in 100 ml dry ether in an ice-salt bath, 2 g anhydrous zinc chloride was added. A steady stream of dry hydrogen chloride gas was passed through the solution for 2 hrs of stirring continuously. The mixture was allowed to stand in refrigerator overnight and again dry hydrogen chloride gas was passed through the mixture for another 2 hours. After keeping the mixture in a refrigerator for three days, the ether was decanted and washed twice with ether; the solid obtained was hydrolyzed by refluxing with 100 ml 2% HCl water for 2 hours. After completion of the reaction the mixture was cooled, filtered, and dried to yield the target compounds.

**1-(2,4,6-Trihydroxy)-2-(4-methoxyphenyl)ethanone (3a).** Yield: 52%; Mp: 190–194°C; Anal. Calcd for C<sub>15</sub>H<sub>14</sub>O<sub>5</sub>: C, 65.69; H, 5.15; O, 29.17; found: C, 65.62; H, 5.19; O, 29.19; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz): δ 12.94 (s, 2 H, -OH), 10.92 (s, 1H, -OH), 7.2 (d, *J* = 8.0 Hz, 2H, Ar-H), 6.8 (d, *J* = 8.0 Hz, 2H,

Ar-H), 5.8 (s, 2H, Ar-H), 4.2 (s, 2H), 3.8 (s, 3H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz): δ 202.6, 164.7, 164.2, 157.8, 130.5, 127.7, 113.5, 103.6, 94.7, 54.9, 47.9; ESI-MS: *m/z* 275 [M + H]<sup>+</sup>.

**1-(2,4,6-Trihydroxyphenyl)-2-phenylethanone (3c).** Yield: 47%, Mp: 158–161°C; Anal. Calcd for C<sub>14</sub>H<sub>12</sub>O<sub>4</sub>: C, 68.85; H, 4.95; O, 26.20; found: C, 68.81; H, 4.98; O, 26.21; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz): δ 12.22 (s, 2H, -OH), 10.39 (s, 1H, -OH), 7.31–7.19 (m, 5H, Ar-H), 5.83 (s, 2H, Ar-H), 4.35 (s, 2H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz): δ 202.2, 164.8, 164.2, 135.9, 129.6, 128.0, 126.1, 103.7, 94.7, 48.8; ESI-MS: *m/z* 245 [M + H]<sup>+</sup>.

**2-(4-Fluorophenyl)-1-(2,4,6-trihydroxyphenyl)ethanone (3d).** Yield: 40%, Mp: 194–196°C; Anal. Calcd for C<sub>14</sub>H<sub>11</sub>FO<sub>4</sub>: C, 64.12; H, 4.23; O, 24.40; found: C, 64.07; H, 4.25; O, 24.43; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz): δ 12.19 (s, 2H, -OH), 10.39 (s, 1H, -OH), 7.27–7.23 (m, 2H, Ar-H), 7.11 (t, *J* = 9.2, 2H, Ar-H), 5.83 (s, 2H, Ar-H), 4.34 (s, 2H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz): δ 202.0, 164.8, 164.2, 160.9 (d, <sup>1</sup>*J*<sub>CF</sub> = 241 Hz, 1C), 132.0 (d, <sup>4</sup>*J*<sub>CF</sub> = 3 Hz, 1C), 131.4 (d, <sup>3</sup>*J*<sub>CF</sub> = 8.0 Hz, 2C), 114.6 (d, <sup>2</sup>*J*<sub>CF</sub> = 21 Hz, 2C), 103.6, 94.7, 48.0; ESI-MS: *m/z* 263 [M + H]<sup>+</sup>.

**1-(2,4,6-Trihydroxyphenyl)-2-(4-nitrophenyl)ethanone (3e).** Yield: 58%, Mp: 222–224°C; Anal. Calcd for C<sub>14</sub>H<sub>11</sub>NO<sub>6</sub>: C, 51.13; H, 3.83; N, 4.84; O, 33.19; found: C, 51.09; H, 3.84; N, 4.85; O, 33.21; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz): δ 12.21 (s, 2H, -OH), 10.92 (s, 1H, -OH), 8.22 (d, *J* = 8.8 Hz, 2H, Ar-H), 7.55 (d, *J* = 8.8 Hz, 2H, Ar-H), 5.89 (s, 2H, Ar-H), 4.57 (s, 2H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz): δ = 200.8, 165.1, 164.2, 146.2, 144.2, 131.1, 123.0, 103.7, 94.7, 48.83; ESI-MS: *m/z* 290 [M + H]<sup>+</sup>.

*1-(2,4,6-Trihydroxyphenyl)-2-p-tolylethanone (3f)*. Yield: 56%, Mp: 168–170°C; Anal. Calcd for  $C_{15}H_{14}O_4$ : C, 69.76; H, 5.46; O, 24.78; found: C, 69.72; H, 5.48; O, 24.80;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.22 (s, 2H, -OH), 10.38 (s, 1H, -OH), 7.09 (s, 4H, Ar-H), 5.81 (s, 2H, Ar-H), 4.28 (s, 2H<sub>3</sub>), 3.01 (s, 3H, CH<sub>3</sub>);  $^{13}C$  NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  202.5, 164.8, 164.2, 135.1, 132.8, 129.4, 128.6, 103.6, 94.6, 48.4, 20.6; ESI-MS:  $m/z$  259 [M + H]<sup>+</sup>.

*2-(4-Bromophenyl)-1-(2,4,6-trihydroxyphenyl)ethanone (3g)*. Yield: 48%, Mp: 218–220°C; Anal. Calcd. for  $C_{14}H_{11}BrO_4$ : C, 52.04; H, 3.43; Br, 24.73; O, 19.80; found: C, 52.01; H, 3.44; O, 19.82;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.12 (s, 2H, -OH), 10.41 (s, 1H, -OH), 7.45 (d,  $J$  = 8.4 Hz, 2H, Ar-H), 7.18 (d,  $J$  = 8.8 Hz, 2H, Ar-H), 5.82 (s, 2H, Ar-H), 4.33 (s, 2H);  $^{13}C$  NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  201.6, 164.9, 164.2, 135.3, 131.9, 130.8, 119.4, 103.6, 94.7, 48.31; ESI-MS:  $m/z$  323 [M + H]<sup>+</sup>.

**3.3.2. General Procedure for the Cyclization of Substituted 2',4',6'-Trihydroxydeoxybenzoins (Synthesis of 4a, 4c–g).** With cooling and vigorous stirring (3 mL, 24 mmol), etherated boron trifluoride was added drop wise to 8 mmol of 3a, 3c–g in 3 ml of anhydrous DMF. The cooling was stopped and phosphorus oxychloride (0.9 ml, 9.6 mmol) was added dropwise; after mixing all the components the reaction mixture was stirred at 60–70°C for 2 h and then poured into acidified water; the precipitate was filtered off and purified by column chromatography using hexane and ethylacetate in the ratio of 8 : 2 as eluents.

*5,7-Dihydroxy-3-(4-methoxyphenyl)-4H-chromen-4-one (4a)*. Yield: 61%, Mp: 212–214°C; Anal. Calcd. for  $C_{16}H_{12}O_5$ : C, 67.60; H, 4.25; O, 28.14; found: C, 67.57; H, 4.27; O, 28.15;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.94 (s, 1H, 5-OH), 10.92 (s, 1H, 7-OH), 8.38 (s, 1H, 2-H), 7.50 (d,  $J$  = 8.0 Hz, 2H, Ar-H), 7.0 (d,  $J$  = 8.0 Hz, 2H, Ar-H), 6.40 (1H, Ar-H), 5.77 (s, 1H, Ar-H), 3.79 (s, 3H);  $^{13}C$  NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  180.0, 164.3, 161.9, 159.17, 157.5, 154.2, 130.1, 122.9, 121.9, 113.7, 104.4, 99.0, 93.6, 55.1; 24; ESI-MS:  $m/z$  285 [M + H]<sup>+</sup>; FT-IR (KBr,  $Cm^{-1}$ ): 3312 (O-H), 3007, (C-H aromatic), 2950 (C-H aliphatic), 1642 (C=O);

*5,7-Dihydroxy-3-phenyl-4H-chromen-4-one (4c)*. Yield: 76%, Mp: 194–196°C; Anal. Calcd. for  $C_{15}H_{10}O_4$ : C, 70.86; H, 3.96; O, 25.17; found: C, 70.82; H, 3.98; O, 25.19;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  10.81 (s, 1H, 7-OH), 8.39 (s, 1H, 2-H), 8.0 (d,  $J$  = 8.8 Hz, 2H, Ar-H), 7.58 (d,  $J$  = 8.0 Hz, 2H, Ar-H), 6.97 (d,  $J$  = 2.0 Hz, 1H, Ar-H), 6.88 (d,  $J$  = 2 Hz, 1H, Ar-H);  $^{13}C$  NMR (DMSO- $d_6$ , 22.5 MHz):  $\delta$  174.3, 162.6, 157.4, 153.6, 153.5, 132.1, 128.8, 127.6, 123.58; ESI-MS:  $m/z$  255 [M + H]<sup>+</sup>; FT-IR (KBr,  $Cm^{-1}$ ): 3198 (O-H), 3062, (C-H aromatic), 2925 (C-Haliphatic), 1626 (C=O).

*3-(4-Fluorophenyl)-5,7-dihydroxy-4H-chromen-4-one (4d)*. Yield: 54%, Mp: 188–190°C; Anal. Calcd. for  $C_{15}H_9FO_4$ : C, 66.18; H, 3.33; F, 6.98; O, 23.51 found: C, 66.13; H, 3.34; O, 23.55;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.83 (s, 1H, 5-OH), 10.93 (s, 1H, 7-OH), 8.44 (s, 1H, 2-H), 7.63 (t,  $J$  = 8.8 Hz, 2H, Ar-H), 7.31 (t,  $J$  = 9.2 Hz, 2H, Ar-H), 6.42 (d,  $J$  = 1.6 Hz, 1H,

Ar-H), 6.25 (d,  $J$  = 1.6 Hz, 1H, Ar-H);  $^{13}C$  NMR (DMSO- $d_6$ , 22.5 MHz):  $\delta$  179.1, 164.6, 161.8, 156.2, 157.3, 147.01, 157.2, 156.2, 147.0, 137.8, 123.10; ESI-MS:  $m/z$  273 [M + H]<sup>+</sup>; FT-IR (KBr,  $Cm^{-1}$ ): 3299 (O-H), 3077, (C-H aromatic), 2924 (C-H aliphatic), 1664 (C=O).

*5,7-Dihydroxy-3-(4-nitrophenyl)-chroman-4-one (4e)*. Yield: 63%, Mp: 286–288°C; Anal. Calcd. for  $C_{15}H_9NO_6$ : C, 60.21; H, 3.03; N, 4.68; O, 32.08; found: C, 60.17; H, 3.05; N, 4.69; O, 32.09;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.70 (s, 1H, 5-OH), 11.01 (s, 1H, 7-OH), 8.63 (s, 1H, 2-H), 8.32 (d,  $J$  = 8.8 Hz, 2H, Ar-H), 7.91 (d,  $J$  = 8.8 Hz, 2H, Ar-H) 6.46 (d,  $J$  = 1.6 Hz, 1H, Ar-H), 6.28 (d,  $J$  = 2 Hz, 1H, Ar-H);  $^{13}C$  NMR (DMSO- $d_6$ , 22.5 MHz):  $\delta$  179.1, 164.6, 161.9, 157.4, 156.1, 146.8, 137.9, 123.15, 120.3, 104.3, 99.2; ESI-MS:  $m/z$  300 [M + H]<sup>+</sup>; FT-IR (KBr,  $Cm^{-1}$ ): 3418 (O-H), 1654 (C=O).

*5,7-Dihydroxy-3-tolyl-4H-chromen-4-one (4f)*. Yield: 68%, Mp: 193–195°C; Anal. Calcd. for  $C_{16}H_{12}O_4$ : C, 71.64; H, 4.51; O, 23.86; found: C, 71.61; H, 4.53; O, 23.87;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.90 (s, 1H, 5-OH), 10.89, (s, 1H, 7-OH), 8.38 (s, 1H, 2-H), 7.46 (d,  $J$  = 8.0 Hz, 2H, Ar-H), 7.26 (d,  $J$  = 8.0 Hz, 2H, Ar-H), 6.40 (d,  $J$  = 2.0 Hz, 1H, Ar-H), 6.24 (d,  $J$  = 2.0 Hz, 1H, Ar-H), 2.34 (s, 3H<sub>3</sub>);  $^{13}C$  NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  179.9, 164.5, 161.99, 157.5, 157.4, 154.5, 137.3, 128.8, 128.7, 127.8, 122.2, 104.4, 99.0, 93.7, 20.7; ESI-MS:  $m/z$  269 [M + H]<sup>+</sup>; FT-IR (KBr,  $Cm^{-1}$ ): 3264 (O-H), 3073, (C-H aromatic), 2921 (C-Haliphatic), 1643 (C=O).

*3-(4-Bromophenyl)-5,7-dihydroxy-4H-chromen-4-one (4g)*. Yield: 66%, Mp: 212–214°C; Anal. Calcd for  $C_{15}H_{14}BrO_5$ : C, 54.08; H, 2.72; Br, 23.99; O, 19.21; found: C, 54.06; H, 2.73; O, 19.22;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.79 (s, 1H, 5-OH), 10.94, (s, 1H, 7-OH), 8.47 (s, 1H, 2-H), 7.66 (d,  $J$  = 8.8 Hz, 2H, Ar-H), 7.55 (d,  $J$  = 8.4 Hz, 2H, Ar-H), 6.42 (d,  $J$  = 1.6 Hz, 1H, Ar-H), 6.25 (d,  $J$  = 1.6 Hz, 1H, Ar-H);  $^{13}C$  NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  179.5, 164.5, 161.9, 155.1, 131.1, 130.9, 121.3, 121.1, 99.2, 93.8; ESI-MS:  $m/z$  332 [M + H]<sup>+</sup>; FT-IR (KBr,  $Cm^{-1}$ ): 3374 (O-H), 3063, (C-H aromatic), 2919 (C-Haliphatic), 1657 (C=O).

**3.3.3. Demethylation of 4a (Preparation of 5,7-Dihydroxy-3-(4-hydroxyphenyl)-4H-chromen-4-one (4b).** 4a (0.26 gm, 0.91 mmol) suspended in toluene (10 ml) was added to anhydrous  $AlCl_3$  and the mixture was refluxed with string to 160°C for 6 h; after completion of reaction (monitored by TLC), the reaction mixture was cooled and poured into water and acidified with HCl acid to break the  $AlCl_3$ ; the solution was then filtered, dried, and washed with toluene and purified by column chromatography to yield 4b.

Yield: 84%, Mp: 292–294°C; Anal. Calcd for  $C_{15}H_{10}O_5$ : C, 66.67; H, 3.73; O, 29.60; found: C, 66.62; H, 3.76; O, 29.62;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  12.94 (s, 1 H, 5-OH), 10.9 (s, 1H, 7-OH), 9.61 (s, 1H, 4'-OH), 8.31 (s, 1H, 2-H), 7.38 (d,  $J$  = 4.0 Hz, 2H, Ar-H), 6.82 (d,  $J$  = 4.0 Hz, 2H, Ar-H), 6.39 (s, 1H, Ar-H);  $^{13}C$  NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  180.2, 164.2, 161.9, 157.5, 157.4, 153.9, 130.1, 122.2, 121.20, 115.0, 104.4, 98.9, 93.6; ESI-MS:  $m/z$  271 [M + H]<sup>+</sup>; FT-IR (KBr,  $Cm^{-1}$ ): 3412 (-OH), 3004, (C-H aromatic), 2933 (C-H aliphatic), 1652 (C=O).



3.3.4. 3-(4-Bromophenyl)-5,7-dimethoxy-4H-chromen-4-one (**4h**) (Prepared by Methylation of **4g**). To a solution of **4g** (6 mmol) in 10 mL acetone, potassium carbonate (18 mmol) was added and then DMS (15 mmol) was added dropwise and refluxed at 60°C for about 2 h. After completion of the reaction as indicated by TLC, the reaction mixture was filtered and concentrated and the crude product was purified by column chromatography (eluent hexane:ethylacetate, 8:2).

Yield: 80%, Mp: 174–176°C; Anal. Calcd for  $C_{17}H_{13}BrO_4$ : C, 56.53; H, 3.63; Br, 22.12; O, 17.72; found: C, 56.50; H, 3.64; O, 17.74;  $^1H$  NMR (DMSO- $d_6$ , 400 MHz):  $\delta$  7.78 (s, 1H, 2-H), 7.52 (d,  $J$  = 8.4 Hz, 2H, Ar-H) 7.43 (d,  $J$  = 8.4 Hz, 2H, Ar-H) 6.45 (d,  $J$  = 2.4 Hz, 1H, Ar-H), 6.38 (d,  $J$  = 2 Hz, 1H, Ar-H), 3.93 (s, 3H), 3.89 (s, 3H);  $^{13}C$  NMR (DMSO- $d_6$ , 100 MHz):  $\delta$  174.8, 164.1, 161.4, 159.8, 150.4, 130.9, 125.3, 122.1, 109.8, 96.3, 96.2, 92.6, 92.5, 56.0; ESI-MS:  $m/z$  361  $[M + H]^+$ ; FT-IR (KBr,  $cm^{-1}$ ): 3002, (C-H aromatic), 2923 (C-H aliphatic), 1657 (C=O).

## Conflicts of Interest

The authors declare that they have no conflicts of interest.

## References

- [1] M. Seeger, M. González, B. Cámara et al., "Biotransformation of natural and synthetic isoflavonoids by two recombinant microbial enzymes," *Applied and Environmental Microbiology*, vol. 38, pp. 473–51, 2003.
- [2] S. A. Fedoreyev, V. P. Bulgakov, O. V. Grishchenko et al., "Isoflavonoid composition of a callus culture of the relict tree *Maackia amurensis* Rupr. et Maxim," *Journal of Agricultural and Food Chemistry*, vol. 56, no. 16, pp. 7023–7031, 2008.
- [3] W. De-Eknamkul, K. Umehara, O. Monthakantirat et al., "QSAR study of natural estrogen-like isoflavonoids and diphenolics from Thai medicinal plants," *Journal of Molecular Graphics and Modelling*, vol. 29, no. 6, pp. 784–794, 2011.
- [4] O. Lapcik, M. Hill, R. Hampl, K. Wähälä, and H. Adlercreutz, "Identification of isoflavonoids in beer," *Steroids*, vol. 63, no. 1, pp. 14–20, 1998.
- [5] G. M. Boland and D. M. X. Donnelly, "Isoflavonoids and related compounds," *Natural Product Reports*, vol. 15, no. 3, pp. 241–260, 1998.
- [6] M. A. Rostagno, M. Palma, and C. G. Barroso, "Pressurized liquid extraction of isoflavones from soybeans," *Analytica Chimica Acta*, vol. 522, no. 2, pp. 169–177, 2004.
- [7] R. Liu, Y. Hu, J. Li, and Z. Lin, "Production of soybean isoflavone genistein in non-legume plants via genetically modified secondary metabolism pathway," *Metabolic Engineering*, pp. 91–79, 2000.
- [8] K. L. Werner, T. Oliver, W. L. Roman, and S. Helga, "Different Types of Combination Effects for the Induction of Micronuclei in Mouse Lymphoma Cells by Binary Mixtures of the Genotoxic Agents MMS, MNU, and Genistein," *Toxicological Sciences*, vol. 86, pp. 318–323, 2005.
- [9] S. Yun-Hee, P. Sun-Dong, and N. J. Kyung-Soo, "Effective chemopreventive activity of genistein against human breast cancer cells," *Biochemistry and Molecular Biology*, vol. 39, p. 451, 2006.
- [10] B. Sanjeev, L. Yiwei, W. Zhiwei, and H. Fazlul, *Cancer Letters*, vol. 269, pp. 226–242, 2008.
- [11] W.-F. Chen, M.-H. Huang, C.-H. Tzang, M. Yang, and M.-S. Wong, "Inhibitory actions of genistein in human breast cancer (MCF-7) cells," *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease*, vol. 1638, no. 2, pp. 187–196, 2003.
- [12] A. Arti, G. Muraleedharan, and M. Gale, "Antioxidant activities of isoflavones and their biological metabolites in a liposomal system," *Free Radical Biology and Medicine*, vol. 24, pp. 1355–1363, 1998.
- [13] C. E. Rüfer and S. E. Kulling, "Antioxidant activity of isoflavones and their major metabolites using different in vitro assays," *Journal of Agricultural and Food Chemistry*, vol. 54, no. 8, pp. 2926–2931, 2006.
- [14] A. Arora, M. G. Nair, and G. M. Strasburg, "Antioxidant activities of isoflavones and their biological metabolites in a liposomal system," *Archives of Biochemistry and Biophysics*, vol. 356, no. 2, pp. 133–141, 1998.
- [15] J. H. Mitchell, P. T. Gardner, D. B. McPhail, P. C. Morrice, A. R. Collins, and G. G. Duthie, "Antioxidant efficacy of phytoestrogens in chemical and biological model systems," *Archives of Biochemistry and Biophysics*, vol. 360, no. 1, pp. 142–148, 1998.
- [16] C. H. Lee, L. Yang, J. Z. Xu, S. Y. V. Yeung, Y. Huang, and Z.-Y. Chen, "Relative antioxidant activity of soybean isoflavones and their glycosides," *Food Chemistry*, vol. 90, no. 4, pp. 735–741, 2005.
- [17] L.-N. Zhang, P. Cao, S.-H. Tan, W. Gu, L. Shi, and H.-L. Zhu, "Synthesis and antimicrobial activities of 7-O-modified genistein derivatives," *European Journal of Medicinal Chemistry*, vol. 43, no. 7, pp. 1543–1551, 2008.
- [18] H.-Q. Li, J.-Y. Xue, L. Shi, S.-Y. Gui, and H.-L. Zhu, "Synthesis, crystal structure and antimicrobial activity of deoxybenzoin derivatives from genistein," *European Journal of Medicinal Chemistry*, vol. 43, no. 3, pp. 662–667, 2008.
- [19] M. Nadal-Serrano, D. Pons, J. Sastre-Serra, and M. Blanquer-Rosselló, "Genistein modulates oxidative stress in breast cancer cell lines according to ER $\alpha$ /ER $\beta$  ratio: Effects on mitochondrial functionality, sirtuins, uncoupling protein 2 and antioxidant enzymes mercedes," *The International Journal of Biochemistry & Cell Biology*, vol. 45, pp. 2051–2051, 2013.
- [20] C. Santell, C. Chang, G. Nair, and G. Helferich, "Dietary genistein exerts estrogenic effects upon the uterus, mammary gland and the hypothalamic/pituitary axis in rats," *Journal of Nutrition*, vol. 127, pp. 263–269, 1997.
- [21] W. H. Tolleson, D. R. Doerge, M. I. Churchwell, M. M. Marques, and D. W. Roberts, "Metabolism of biochanin A and formononetin by human liver microsomes in vitro," *Journal of Agricultural and Food Chemistry*, vol. 50, no. 17, pp. 4783–4790, 2002.
- [22] L. Kole, B. Giri, S. K. Manna, B. Pal, and S. Ghosh, "Biochanin-A, an isoflavon, showed anti-proliferative and anti-inflammatory activities through the inhibition of iNOS expression, p38-MAPK and ATF-2 phosphorylation and blocking NF $\kappa$ B nuclear translocation," *European Journal of Pharmacology*, vol. 653, no. 1–3, pp. 8–15, 2011.
- [23] Y. J. Seo, B. S. Kim, S. Y. Chun, Y. K. Park, K. S. Kang, and T. G. Kwon, "Apoptotic effects of genistein, biochanin-A and apigenin on LNCaP and PC-3 cells by p21 through transcriptional inhibition of polo-like kinase-1," *Journal of Korean Medical Science*, vol. 26, no. 11, pp. 1489–1494, 2011.
- [24] K. Polkowski and A. P. Mazurek, "Biological properties of genistein. A review of in vitro and in vivo data," *Acta Poloniae Pharmaceutica. Drug Research*, vol. 57, no. 2, pp. 135–155, 2000.

- [25] M. Switalska, G. Gryniewicz, L. Strzadala, and J. Wietrzyk, "Novel genistein derivatives induce cell death and cell cycle arrest through different mechanisms," *Nutrition and Cancer*, vol. 65, no. 6, pp. 874–884, 2013.
- [26] Y.-W. Kim, J. C. Hackett, and R. W. Brueggemeier, "Synthesis and aromatase inhibitory activity of novel pyridine-containing isoflavones," *Journal of Medicinal Chemistry*, vol. 47, no. 16, pp. 4032–4040, 2004.
- [27] M. Radharani, A. Madhan, A. Ravi, S. Vered, B. U. Christopher, and K. Saeed, *Cancer Biology & Therapy*, vol. 11, pp. 883–892, 2011.
- [28] K. V. Raghava Rao and T. Raghava Rao, "Molecular characterization and its antioxidant activity of a newly isolated *Streptomyces coelicoflavus* BC 01 from mangrove soil," *Journal of Young Pharmacists*, vol. 5, no. 4, pp. 121–126, 2013.
- [29] B. Frédéric, D. Aline, B. AHCÈME et al., "Genistein and fluorinated analogs suppress agonist-induced airway smooth muscle contraction," *Bioorganic & Medicinal Chemistry Letters*, vol. 7, pp. 1323–1326, 1997.

